

FRACTIONATED SOYBEAN PROTEIN AND
PROCESS FOR PRODUCING THE SAME

5 Technical Field

The present invention relates to a process for producing a 7S globulin-rich fraction and an 11S globulin-rich fraction from a solution containing soybean protein, and the soybean protein obtained from the process.

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Background Art

Soybean storage protein is precipitated at about pH 4.5 and can be relatively easily separated from components other than the protein. This is referred to as a soybean protein isolate and, in many cases, soybean protein in this form is utilized in the food industry. The soybean storage protein is further divided into 2S, 7S, 11S and 15S globulins according to sedimentation constants in ultracentrifugation analysis. Among them, 7S globulin and 11S globulin are predominant constituent protein components of the globulin fractions (note: 7S globulin and 11S globulin are classification names in a sedimentation method and substantially correspond to β -conglycinin and glycinin according to immunological nomenclature, respectively), and both of them have specific different properties such as

viscosity, coagulability, surface activity, etc. Then, fractionation of soybean protein into 7S globulin-rich fraction and 11S globulin-rich fraction makes it possible to utilize properties of respective protein components, and 5 it is expected to expand industrial utilization of proteins.

7S Globulin and 11S globulin are composed of several subunits. 7S Globulin is composed of three subunits, i.e., α , α' and β subunits. 11S Globulin is composed of a pair of an acidic polypeptide (A) and a basic polypeptide (B) each of which has several subunits. The composition ratio of these subunits in most common conventional soybeans is typically about 1 : 2 in the ratio of 7S globulin to 11S globulin as determined from the densitometry area ratio of migration patterns obtained by SDS-polyacrylamide gel 10 electrophoresis. The properties of 7S globulin and 11S globulin resemble in their molecular weights and charge states. In particular, both globulins are diversified due to combinations of subunits, and properties thereof range to some extent to thereby overlap each other. Accordingly, 15 it is not easy to efficiently separate these globulins with little contamination to one another. 20

Conventionally known fractionation methods are as follows. That is, these methods include a method utilizing a difference in isoelectric points: only 7S globulin is 25 extracted in the vicinity of the isoelectric point of 11S

globulin (JP 55-124457 A); a method utilizing a difference in reactivity with a calcium salt: a 7S globulin-rich fraction is extracted by addition of a small amount of calcium during extraction (JP-A 48-56843 A); a method utilizing a difference in solubility at a certain pH and 5 ionic strength: 7S globulin is prepared by removing insoluble fractions in the presence of sodium chloride or potassium chloride in a pH region of 1.2 to 4.0 (JP 49-31843 A), 7S globulin and 11S globulin are separated by 10 adjusting pH of a slurry after isoelectric point precipitation to 5.0 to 5.6 while adjusting sodium chloride concentration to 0.01 to 0.2 M (JP 58-36345 A), and 7S globulin is fractionated by adjusting the ionic strength of 15 a protein-containing solution to 0.2 to 0.3 and the pH to 4.8 to 5.2 to remove insoluble fractions, and then adjusting the ionic strength to less than 0.2 and the pH to 4.6 to 5.0 (JP 5-43597 A); and a method utilizing a cold-precipitation phenomenon and a reducing agent, etc.: this utilizes a phenomenon that the solubility of 11S globulin 20 is lowered at a low temperature (referred to as cryo-precipitation phenomenon), and a soybean protein raw material is treated in the presence of a sulfurous acid compound, glutathione compound or cysteine compound in an aqueous system at pH 6.5 or higher, followed by adjusting 25 pH to 5.5 to 7.0 and a temperature to 22°C or lower to

fractionate into a 7S globulin-rich soluble fraction and an 11S globulin-rich insoluble fraction (JP 61-187755 A).

These known fractionation methods skillfully utilize a difference in solubility between 7S globulin and 11S globulin due to pH, ionic strength, the presence of a certain salt, temperature, etc. However, there are such problems that these known fractionation methods are unsuitable for an industrially applicable fractionation method because, for example, separation by a high centrifugal force is required for clear fractionation. Thus, problems still remain in practice. For example, in the method of JP 61-187755 A, cryo-precipitation phenomenon highly depends on a temperature and it is necessary to cool a reaction mixture to about 5°C, which results in such a practical problem that a large amount of a sulfurous acid compound, etc. should be added to separate fractions with an industrially available low centrifugal force, as well as which results in such a problem of fractionation precision that a little amount of 11S globulin is contaminated in a soluble fraction.

For the purpose of obtaining 7S globulin-rich protein, isolation of protein from 11S globulin-defect soybeans, i.e., a 7S globulin-rich seeds produced by breeding has been studied (Breeding Science, 46, 11, 1996), and its utility (Breeding Science, 50, 101, 2000) and patent (US

6,171,640 B1) can also be found.

As described above, a method for separating a 7S globulin-rich fraction and a 11S globulin-rich fraction by which an mutual contamination between soluble and insoluble fractions is reduced and by which a production in an industrial scale is accomplished conveniently and efficiently has been researched and developed.
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On the other hand, it has been reported by Samoto et al. that, in soybean-derived protein, there is a protein component having high affinities with polar lipid as a constituent of a cytoplasmic membrane as well as a protein body or oil body membrane (oil-body-associated protein) which amounts to as high as about 35 % of an industrially produced soybean protein isolate (Biosci. Biotechnol. 10 Biochem., 62 (5), 935-940 (1998)). The oil-body-associated protein is a general term of protein components consisting mainly of membrane protein, especially those whose molecular weights measured by SDS-polyacrylamide gel electrophoresis are mainly 34 kDa, 24 kDa and 18 kDa, and 15 contains about 10 to 12% by weight of polar lipid which are extractable with a 2 : 1 polar solvent mixture of chloroform : ethanol.
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Conventional fractionation is focusing only on 7S and 11S globulins, and in many cases, pays no attention to the 25 oil-body-associated protein which is contaminating each

fraction, since the oil-body-associated protein can not be identified as positively as 7S and the 11S globulins when analyzed by SDS-polyacrylamide gel electrophoresis and is underestimated frequently. In other words, a purity when determined only by SDS polyacrylamide gel electrophoresis becomes higher frequently than the actual purity, and the behavior of the oil-body-associated protein should be taken into account for the purpose of obtaining 7S or 11S globulin at a really high purity. Thus, a conventional fractionation into the two fractions 7S globulin/11S globulin handles a purity of a fraction only as a ratio between 7S globulin and 11S globulin. However, each fraction is associated with the oil-body-associated protein, and in many cases its actual state is a relatively crude fraction having a slightly lower purity whose protein composition is characterized by a large amount of the oil-body-associated protein.

While the present inventors have proposed a method which makes industrial fractionation of 7S globulin and 11S globulin possible only by heat-treating under acidic conditions (WO 02/28198 A1), as a result of an intensive study, it has been found that pH for separating a soluble fraction containing 7S globulin from an insoluble fraction containing 11S globulin can be lowered by combining heat treatment of a solution containing soybean protein under

acidic conditions with adjustment of an ionic strength, thereby further facilitating separation of the soluble fraction and the insoluble fraction.

The present invention proposes a novel fractionation method between 7S globulin and 11S globulin, in particular, one object of the present invention is a highly accurate and efficient fractionation method which can be performed in an industrial scale. Another object of the present invention is to provide protein fractions having characteristic features with less contamination of the oil-body-associated protein and having high purities of 7S globulin and 11S globulin.

10 Patent Document 1: JP 55-124457 A

15 Patent Document 2: JP 48-56843 A

Patent Document 3: JP 49-31843 A

Patent Document 4: JP 58-36345 A

Patent Document 5: JP 5-43597 A

Patent Document 6: JP 61-187755 A

Patent Document 7: US 6,171,640 B1

20 Patent Document 8: WO 02/28198 A1

Non-patent Document 1: K. Yagasaki etc., Breeding Science, 46, 11, 1996

25 Non-patent Document 2: K. Yagasaki etc., Breeding Science, 50, 101, 2000

Non-patent Document 3: M. Samoto etc., Biosci.

Biotechnol. Biochem., 62(5), 935-940, 1998

Disclosure of the Invention

The present invention relates to:

- 5 (1) A process for producing soybean protein, which is comprises heating a solution containing the soybean protein under acidic conditions, and then fractionating it into a soluble fraction and an insoluble fraction at an ionic strength of 0.02 or more and pH of 4.5 or higher but lower
10 than 5.6;
- 15 (2) The process according to (1), wherein the solution containing the soybean protein is an aqueous slurry of defatted soybeans, defatted soybean milk obtained from the slurry, a slurry of acid-precipitated soybean protein, or a solution of soybean protein isolate;
- 15 (3) The process according to (1), wherein the acidic conditions are those at pH 3.8 to 6.8;
- 15 (4) The process according to (1), wherein the heating is performed at 30 to 75°C;
- 20 (5) The process according to (1), which further comprises fractionating 7S globulin protein from the soluble fraction obtained by the fractionation in (1), wherein a ratio of 7S globulin/(11S globulin + 7S globulin) of said 7S globulin protein is 0.5 or more, and a content
25 of a polar lipid extracted by a mixed solvent of chloroform

and methanol (chloroform : methanol = 2 : 1) in the solid content of said 7S globulin protein is 1% by weight or less;

(6) 7S Globulin protein obtained by the process
5 according to (5), whose phytic acid content is 1.2% by weight or less in the solid content;

(7) The process according to (1), which further comprises fractionating 11S globulin protein from the insoluble fraction obtained by the fractionation in (1),
10 wherein a ratio of 11S globulin/(11S globulin + 7S globulin) of said 11S globulin protein is 0.7 or more, and a content of a polar lipid extracted with a mixed solvent of chloroform and methanol (chloroform : methanol = 2 : 1) in the solid content of said 11S globulin protein is 2% by
15 weight or less;

(8) 11S Globulin protein obtained by the process according to (7), whose phytic acid content is 1.2% by weight or less in the solid content.

20 Best Mode for Performing the Invention

The method of the present invention is characterized in that, in comparison with the method which makes industrial fractionation of 7S globulin and 11S globulin possible only by heat-treating under acidic conditions (WO
25 02/28198 A1), pH for separating a soluble fraction

containing 7S globulin from an insoluble fraction containing 11S globulin can be lowered by combining heat treatment of a solution containing soybean protein under acidic conditions with adjustment of an ionic strength,
5 thereby further facilitating separation of the soluble fraction and the insoluble fraction. The above acidic conditions are preferably those at pH 3.8 to 6.8, the heating temperature is preferably 30 to 75°C, the ionic strength is preferably 0.02 or more, and the separation of
10 the soluble fraction from the insoluble fraction is preferably performed at a pH of 4.5 or more but lower than 5.6. Whereby a 7S globulin-rich soluble fraction containing less oil-body-associated protein can be obtained, and further, an 11S globulin and oil-body-associated
15 protein-rich insoluble fraction can be obtained. It is possible to selectively dissolve and separate 11S globulin with keeping the oil-body-associated protein insoluble by applying weak shear stress to the above insoluble fraction in an aqueous solution at an approximately neutral pH (pH
20 6.5 to 7.5), thereby dissolving or extracting 11S globulin in the insoluble fraction to obtain a 11S globulin-rich fraction containing less oil-body-associated protein.

Further, in the present invention, a protein fraction with a low phytic acid content of 1.2% or less of phytic
25 acid per protein can be obtained by decomposing phytic acid

by a phytase during the production step. The separation efficiency can be further improved by performing the phytase treatment before separation of the soluble fraction and the insoluble fraction.

5 The soluble fraction obtained by the above production process has a ratio of 7S globulin/(7S globulin + 11S globulin) of 0.5 or more, and a high purified 7S globulin fraction with the above ratio of 0.8 or more, 0.85 or more, or 0.9 or more can be readily obtained by appropriately
10 selecting pH for separating the soluble fraction and the insoluble fraction. Another fraction, i.e., the insoluble fraction, has a 11S globulin/(11S globulin + 7S globulin) ratio of 0.7 or more, and a high purified 11S globulin fraction with the above ratio of 0.8 or more, 0.85 or more,
15 or 0.90 or more can be readily obtained by appropriately selecting the pH for separating the soluble fraction and the insoluble fraction.

 Herein, the ratio of 7S globulin/(11S globulin + 7S globulin) or 11S globulin/(11S globulin + 7S globulin) can
20 be determined from the area ratio between the corresponding fractions by measuring the migration pattern obtained by SDS-polyacrylamide electrophoresis with a densitometry.

 The oil-body-associated protein is present in a proportion of about 30 to 35% in acid-precipitated protein,
25 when the protein is obtained precisely using sodium sulfate

from the acid-precipitated protein that is substantially free from denaturation (Biosci. Biotechnol. Biochem., 62(5), 935-940 (1988)). In view of the fact that polar lipid extracted by a mixed solvent of chloroform and methanol in 5 a ratio of 2 : 1 (volume ratio) is present in a proportion of 3 to 4% by weight in the solid content of the acid-precipitated protein and in a proportion of 10 to 12% by weight in the oil-body-associated protein, the above polar lipid (hereinafter, sometimes, abbreviated as "chloroform-methanol oil content") is unevenly distributed in the oil-body-associated protein of the acid-precipitated protein, and the amount of the oil-body-associated protein can be calculated to be 10 fold by weight of the chloroform-methanol oil content. However, this calculation is 10 applicable to a subject material prepared via a defatting step with hexane, etc., and in case of a subject material which is not subjected to hexane extraction, the 15 calculation is firstly applicable after defatting with hexane.

20 The 7S globulin-rich soluble fraction obtained in the present invention contains 1% or less of the chloroform-methanol oil content, and the content of the oil-body-associated protein is estimated to be 10% or less. The 11S globulin-rich fraction extracted from the insoluble 25 fraction contains 2% or less of the chloroform-methanol oil

content, and the content of the oil-body-associated protein is calculated to be 20% or less.

The assay method used in the present invention will be described below.

5 * Crude protein: The nitrogen content was determined by Kjeldahl method, and it was multiplied by a coefficient of 6.25 to convert it into the amount of crude protein.

10 * Separation-precipitation rate of insoluble fraction: A separation characteristic analyzer LUMiFuge 114 manufactured by L.U.M. Co. was used. A sample solution (0.5 ml) containing the insoluble fraction was placed in a polystyrene rectangular cell, the cell was centrifuged at 100G, and the moving rate of the interface with a transmittance of 20% was defined as the separation-
15 precipitation rate.

20 * SDS-polyacrylamide electrophoresis: A gradient gel with a gel concentration of 10% to 20% according to the method by Laemmli (Nature, 227, 680 (1970)) was used to analyze protein. The applied amount of the protein was 6 μ g in terms of a solid content, and the gel was stained with Coomassie Brilliant Blue R-250.

* Phytic acid: The amount of phytic acid was assayed according to Alii Mohamed method (Cereal Chemistry 63, 475-478 (1986)).

25 * Chloroform-methanol oil content: About 50 fold in

volume of a mixed solution of chloroform-methanol (volume ratio, 2 : 1) was added to a dried sample, and the weight ratio of the solid content extracted by refluxing the solvent was measured as the chloroform-methanol oil content.

5 * Purity (SPE standard): The migration pattern obtained by the above SDS-polyacrylamide electrophoresis was measured with a densitometry. The area ratio of the desired fraction relative to the total area of the pattern was defined to be the purity (SPE standard). The content 10 of 7S globulin refers to the total content of α , α' and β subunits, and the content of 11S globulin refers to the total content of the acidic polypeptide (A) and basic polypeptide (B).

15 * Corrected purity: The corrected purity was calculated from the above-obtained purity (SPE standard) as follows by taking the oil-body-associated protein contaminated therein into consideration. That is, since a sample contains the oil-body-associated protein corresponding to 10 fold by weight of the chloroform-methanol oil content in addition to 7S globulin and 11S globulin, assuming A (%) represents the purity (SPE standard), a corrected purity is calculated relative to the total protein including 7S globulin, 11S globulin and the oil-body-associated protein:

25 Corrected purity (%) = (100 (%)) - chloroform-methanol

oil content (%) × 10) × A (%) / 100

* Ionic strength: Conductivity of a solution was measured using a conductivity meter (2%/°C, with a temperature conversion function), and a molar concentration of NaCl corresponding to the measured conductivity was defined to be the ionic strength.

Preferred embodiments of the present invention will be described below.

As raw material soybeans used in the present invention, any soybeans of commercially available soybeans, or soybeans deficient in a specified fraction produced by bleeding or genetic engineering can be used. While the solution containing soybean protein may be any of an aqueous slurry of defatted soybeans (hereinafter referred to as a defatted soybean slurry), defatted soybean milk obtained from the slurry, a slurry of acid-precipitated soybean protein (hereinafter referred to as a curd slurry), or a solution of a soybean protein isolate, in particular, to use an aqueous slurry of defatted soybeans is preferable since the soluble fraction and the insoluble fraction are readily separated. A solution of non-denatured or scarcely denatured soybean protein is preferable in order to fractionate into a 7S globulin-rich fraction and an 11S globulin-rich fraction.

The heating of the solution containing soybean protein

under acidic conditions is performed at pH 3.8 to 6.8, more preferably pH 4.0 to 6.6, further preferably pH 4.2 to 6.2 at a temperature of 30 to 75°C, more preferably 35 to 65°C, and further preferably 40 to 60°C. The ionic strength of 5 0.02 or more facilitates separation of the soluble fraction and the insoluble fraction, and the higher ionic strength can further facilitate separation of the soluble fraction and the insoluble fraction. However, since an ionic strength should be adjusted to less than 0.2 for 10 isoelectric point precipitation of 7S globulin from the soluble fraction after separation when the ionic strength of 0.2 or more is used for separating the fraction, an ionic strength of 0.02 or more but less than 0.2 is recommended for avoiding complex separation procedures.

15 The 7S globulin-rich soluble fraction and the 11S globulin-rich insoluble fraction can be obtained when the soluble fraction is separated from the insoluble fraction at pH of 4.5 or more but less than 5.6. Separation of the 7S globulin-rich fraction and the 11S globulin-rich fraction 20 can be further facilitated by decomposing phytic acid contained in the soybean protein with phytase during the production process, particularly in any steps before the soluble fraction and the insoluble fraction are separated. The phytase treatment is simplified and facilitated when it 25 is performed simultaneously with the heat treatment under

acidic conditions. The optimum conditions for the phytase treatment are usually at pH of 3.5 to 9.0, and a temperature of 20 to 70°C for 5 minutes to 3 hours with about 0.1 to 100 units of a phytase per protein weight (g),
5 though the conditions somewhat may vary depending on the origin of the phytase. The phytase activity of 1 unit is defined by the amount of the enzyme required for releasing 1 μ mole of phosphoric acid from phytic acid as the substrate in 1 minute at the initial stage of the enzymatic
10 reaction at pH 5.5 and 37°C.

Unlike to the pH and the temperature in the heat treatment described above, the length of the duration of the heating has no serious effect on the difficulty experienced in a separation process, and thus is of a less
15 significance. A pH departing out of the range from pH 3.8 to 6.8 or a temperature out of the range from 30°C to 75°C in the heating treatment may lead to a difficulty in separating into 7S globulin and 11S globulin.

After the heating treatment, the fractionation may be
20 performed at the same temperature, although it is preferable to ensure a cooling for controlling any microbes. The fractionation may be accomplished by using a known separation procedure (such as filtration and centrifugation), and an easy separation can also be
25 accomplished especially by using a continuous centrifuge

(such as a decanter). It is a matter of course that the use of a non-continuous centrifuge such as a batch centrifuge is not excluded.

The accuracy of fractionation into the 7S globulin-rich soluble fraction and the 11S globulin-rich insoluble fraction according to the present invention can be evaluated based on the pattern obtained by SDS-polyacrylamide electrophoresis (the purity is in SPE standard).

However, since the purity based on SPE standard may be underestimated due to low stainability of the oil-body-associated protein on the SDS-polyacrylamide electrophoresis, the corrected purity obtained by the above equation:

Corrected purity (%) = (100 (%) - chloroform-methanol oil content (%) × 10) × A (%) / 100
is considered to be closer to the true purity.

After separation, the soluble fraction can be used as the 7S globulin-rich fraction as such or after concentration, neutralization, sterilization or drying.

The concentration can be performed, for example, by adjusting the ionic strength at less than 0.2 with pH adjustment of 4.0 to 5.0 to separate and recover the resulting insoluble fraction. Usually, the soluble fraction is further diluted with water, neutralized,

sterilized by heating, and dried. The heat sterilization can be performed by a known method such as HTST, UHT treatment. Of course, the fraction may be treated with an enzyme such as a protease in a solution state according to
5 a particular purpose.

11S Globulin can be dissolved or extracted from the insoluble fraction after separation using an appropriately neutral aqueous solution (at about pH 6.5 to 7.5) in order to separate 11S globulin from the insoluble oil-body-associated protein [including "okara (insoluble residue from soybean milk or tofu production)" component when the insoluble fraction contains it]. At this time, it is preferable to dissolve or extract 11S globulin by applying shear stress as weak as possible in order to selectively
10 dissolve or extract 11S globulin while preventing solubilization of the oil-body-associated protein fraction. A high-G centrifuge at centrifugal force of about 4,000 G or more, preferably about 5,000 G or more, is favorably used for separating extracted or dissolved 11S globulin
15 from the oil-body-associated protein, thereby making it possible to obtain a protein fraction with a chloroform-methanol oil content of 2% or less in the solid content while maintaining the proportion of 11S globulin of 0.7 or more relative to the total amount of 11S globulin and 7S
20 globulin.
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The 11S globulin-rich fraction, which is dissolved or extracted using an approximately neutral aqueous solution and contains 2% or less of the extracted chloroform-methanol oil content, can be used as the 11S globulin-rich fraction as such or after concentration, neutralization, sterilization or drying. The concentration can be performed, for example, by adjusting the soluble fraction at pH 4.5 or more but less than 5.6 to separate and recover the resulting insoluble fraction. The insoluble fraction is usually further diluted with water, neutralized and sterilized by heating followed by drying. The heat sterilization is usually performed by a known method such as HTST and UHT treatment. Of course, the insoluble fraction may be treated with an enzyme such as a protease in a solution state according to a particular purpose.

The following Examples will further illustrate the present invention, but are not to be construed to limit technical scope of the present invention.

Example 1

To scarcely denatured and defatted soybeans (1 part by weight, nitrogen solubilization index (NSI): 91), which was obtained by flaking soybeans and extracting their oil with an extraction solvent, n-hexane, to separate and remove the oil, was added extraction water (10 parts by weight, ion-exchange water). The mixture was stirred with a homo-mixer

at 22°C and extracted for 40 minutes with maintaining at pH 7.2 by addition of a 20% sodium hydroxide solution. Then, the extract was filtered by a filter cloth, and centrifuged at 5,000 G for 10 minutes to remove insolubles, thereby

5 obtaining defatted-soybean milk. The soybean milk was adjusted to pH 4.5 with 35% hydrochloric acid, and centrifuged at 3,000 G for 10 minutes to obtain acid-precipitated protein. Ion-exchange water was added to the acid-precipitated protein so that the protein content was

10 5% as dry weight, and the solution was homogenized with Polytron (manufactured by KINEMATICA AG) (hereinafter referred to as a curd slurry). The ionic strength of the curd slurry was adjusted to 0.14 with sodium chloride, and stirred at 22°C for 30 minutes, and the pH of the curd

15 slurry was adjusted to 5.3 with a 20% aqueous sodium hydroxide solution, followed by stirring at 22°C for 20 minutes. Then, the curd slurry was heated to 50°C and, after stirring at 50°C for 10 minutes, the curd slurry was immediately cooled to 22°C. The time required from the

20 start of heating to cooling to 22°C was 25 minutes. After further stirring at 22°C for 15 minutes, the separation-precipitation rate of the insoluble fraction was measured using LUMiFuge 114. At the same time, the soluble fraction and insoluble fraction obtained by centrifugation at 5,000

25 G for 5 minutes were assayed by SDS-polyacrylamide

electrophoresis, and the ratios of 7S globulin and 11S globulin (the fraction area ratio obtained by measuring migration patterns of SDS-polyacrylamide electrophoresis with a densitometer) were determined.

5 The separation-precipitation rate of the insoluble fraction is shown in Table 1, and the ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction are shown in Table 2.

Comparative Example 1

10 The ionic strength of a 5% curd slurry prepared according to the same manner as in Example 1 was adjusted to 0.14 with sodium chloride. The curd slurry was stirred at 22°C for 30 minutes, adjusted to pH 5.3 with a 20% aqueous sodium hydroxide solution, and then stirred at 22°C
15 for 60 minutes. According to the same manner as in Example 1, the separation-precipitation rate of the insoluble fraction was measured, and the soluble fraction and insoluble fraction obtained by centrifugation at 5,000 G for 10 minutes were assayed by SDS-polyacrylamide
20 electrophoresis to determine the ratios of 7S globulin and 11S globulin.

25 The separation-precipitation rate of the insoluble fraction is shown in Table 1, and the ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction are shown in Table 2.

Table 1

Separation-precipitation rate of insoluble fraction

	With heating	Without heating
Separation-precipitation rate ($\mu\text{m/sec}$)	146	57

Table 2

Ratio of 7S globulin and 11S globulin (7S:11S)

	Soluble fraction	Insoluble fraction
With heating	98:2	15:85
Without heating	98:2	13:87

As seen from the results in Example 1 and Comparative Example 1, it is clear that separation of the soluble fraction containing highly purified 7S globulin and the insoluble fraction containing highly purified 11S globulin can be further facilitated by heating the curd slurry under acidic conditions.

Example 2

The ionic strength of a 5% curd slurry prepared according to the same manner as in Example 1 was adjusted to 0.14 with sodium chloride. The curd slurry was stirred at 22°C for 30 minutes without adjusting pH (about pH 4.5), and heated to 50°C, followed by immediately cooling to 22°C. The time required from the start of heating to cooling to 22°C was 15 minutes. After cooling, pH of the curd slurry was adjusted to 5.5 with a 20% aqueous sodium hydroxide

solution followed by stirring for 15 minutes. The separation-precipitation rate of the insoluble fraction was measured according to the same manner as in Example 1, and the ratios of 7S globulin and 11S globulin in the soluble 5 fraction and insoluble fraction were determined.

The ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction are shown in Table 3, and the separation-precipitation rate of the insoluble fraction is shown in Table 4.

10 Comparative Example 2

A 5% curd slurry prepared according to the same manner as in Example 1 was subjected to heat treatment according to the same manner as in Example 2 (pH was not adjusted; the slurry was stirred at 22°C for 30 minutes, and heated 15 to 50°C followed by immediate cooling to 22°C) without adjusting the ionic strength (ionic strength: 0.013). Then, pH of the curd slurry was adjusted to 5.9 with a 20% aqueous sodium hydroxide solution (a fraction having a similar ratio of 7S globulin and 11S globulin was not able 20 to be obtained unless pH for separating the soluble fraction and the insoluble fraction was raised when the ionic strength was low). After stirring for 15 minutes, the separation-precipitation rate of the insoluble fraction was measured according to the same manner as in Example 1, 25 and the ratios of 7S globulin and 11S globulin in the

soluble fraction and insoluble fraction were measured.

The ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction are shown in Table 3, and the separation-precipitation rate of the insoluble

5 fraction is shown in Table 4.

Table 3

The ratio of 7S globulin and 11S globulin (7S : 11S)

Ionic strength	Separation pH	Soluble fraction	Insoluble fraction
0.14	5.5	84:16	18:82
0.013	5.9	82:18	15:85

Table 4

Separation-precipitation ratio of insoluble fraction

Ionic strength	0.14	0.013
Separation-precipitation rate ($\mu\text{m/sec}$)	204	48

As seen from the results in Example 2 and Comparative Example 2, it is clear that separation of the soluble fraction containing 7S globulin from the insoluble fraction containing 11S globulin can be further facilitated by raising the ionic strength and lowering the separation pH.

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As seen from the results in the above Examples and Comparative Examples, it is clear that separation of the soluble fraction containing 7S globulin and the insoluble fraction containing 11S globulin is further facilitated by combining heat treatment of a soybean protein containing

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solution under acidic conditions with adjustment of the ionic strength, as compared with the heat treatment only under acidic conditions or adjustment of the ionic strength only.

5 Example 3

Extraction water (ion-exchange water, 10 parts by weight, 22°C) was added to 1 part by weight of scarcely denatured and defatted soybeans defatted according to the same manner as in Example 1 (hereinafter referred to as a 10 defatted soybean slurry). The ionic strength was adjusted to 0.17 with sodium chloride, and the curd slurry was stirred with a propeller at 22°C for 30 minutes without adjusting the pH. Then, pH was adjusted to 5.3 with 35% hydrochloric acid, and the crude slurry was heated to 50°C, 15 and stirred with the propeller for 10 minutes, followed by immediate cooling to 22°C. The time required for heating to 50°C was 10 minutes, while the time required for cooling to 22°C was 5 minutes. After cooling, the crude slurry was adjusted to pH 4.8 with 35% hydrochloric acid and, after 20 stirring at 22°C for additional 10 minutes, the separation-precipitation rate of the insoluble fraction was measured according to the same manner as in Example 1. Further, the ratio of 7S globulin and 11S globulin was determined according to the same manner as in Example 1 with respect 25 to the soluble fraction obtained by centrifugation at 5,000

G for 5 minutes. On the other hand, water (7 fold by weight of defatted soybeans) was added to the insoluble fraction, and protein was extracted at 22°C by stirring with a homo-mixer for 30 minutes while the pH was maintained at 7.2 by adding a 20% aqueous sodium hydroxide solution, and the soluble fraction obtained by centrifugation at 5,000 G for 5 minutes was assayed by SDS-polyacrylamide electrophoresis to determine the ratio of 7S globulin and 11S globulin.

The ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction, respectively, are shown in Table 5, and the separation-precipitation rate of the insoluble fraction is shown in Table 6.

Comparative Example 3

The ionic strength of a defatted soybean slurry prepared according to the same manner as in Example 3 was adjusted to 0.17 with sodium chloride, and the slurry was stirred at 22°C for 30 minutes. Then, pH of the defatted soybean slurry was adjusted to 5.3 with 35% hydrochloric acid, followed by stirring at 22°C for 25 minutes. Then, pH of the slurry was adjusted to 4.8 with a 20% aqueous sodium hydroxide solution and, after stirring at 22°C for 10 minutes, the separation-precipitation rate of the insoluble fraction was measured, and the ratios of 7S globulin and 11S globulin in the soluble fraction and

insoluble fraction were measured according to the same manner as Example 1.

The ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction are shown in Table 5, and the separation-precipitation rate of the insoluble fraction is shown in Table 6.

Table 5

Ratio between 7S globulin and 11S globulin (7S:11S)

	Soluble fraction	Insoluble fraction
With heating	92:8	9:91
Without heating	91:9	10:90

Table 6

Separation-precipitation rate of insoluble fraction

	With heating	Without heating
Separation-precipitation rate ($\mu\text{m/sec}$)	395	84

As seen from the results in Example 3 and Comparative Example 3, it is clear that separation of the soluble fraction and the insoluble fraction is also facilitated by adjusting the heat treatment temperature and ionic strength under acidic conditions when the defatted soybean slurry is used.

Example 4

The ionic strength of a 5% curd slurry prepared according to the same manner as in Example 1 was adjusted

to 0.17 with sodium chloride. After heating the slurry according to the same manner as in Example 3 (the slurry was stirred at 22°C for 30 minutes with a propeller without adjusting pH, pH was adjusted to 5.3, and the slurry was 5 heated to 50°C, stirred with a propeller followed by immediate cooling to 22°C). Then, pH was adjusted to 5.4 with a 20% aqueous sodium hydroxide solution (pH for separation of the curd slurry should be higher than that for separation of the defatted soybean slurry for obtaining 10 fractions with the same ratios of 7S globulin and 11S globulin from the defatted soybean slurry and curd slurry, when the ionic strength of the defatted soybean slurry was the same as that of the curd slurry) followed by stirring at 22°C for additional 10 minutes. The separation- 15 precipitation rate of the insoluble fraction was measured according to the same manner as in Example 1, and the ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction were determined by SDS-polyacrylamide electrophoresis.

20 The ratios of 7S globulin and 11S globulin in the soluble fraction and insoluble fraction are shown in Table 7, and the separation-precipitation rate of the insoluble fraction is shown in Table 8.

Table 7

Ratio between 7S globulin and 11S globulin (7S:11S)

	Soluble fraction	Insoluble fraction
Curd slurry	93:7	9:91

Table 8

Separation-precipitation rate of insoluble fraction

	Curd slurry
Separation-precipitation rate ($\mu\text{m/sec}$)	156

As seen from the results in Example 3 and Example 4, it is clear that separation is much facilitated with a higher separation-precipitation rate of the insoluble fraction in case of using the defatted soybean slurry in the solution containing soybean protein.

Example 5

The ionic strength of a 5% curd slurry prepared by the same method as in Example 1 was adjusted to 0:14 with sodium chloride, and the slurry was stirred at 22°C for 30 minutes without adjusting the pH (about pH 4.5) followed by heating to 50°C. The time required for raising the temperature to 50°C was 10 minutes. After the temperature was raised to 50°C, pH of the slurry was adjusted to 5.3 with a 20% aqueous sodium hydroxide solution, and the slurry was heated for additional 20 minutes followed by cooling to 22°C. The time required for cooling was 5 minutes. After cooling, the slurry was centrifuged at

5,000 G for 10 minutes to obtain the soluble fraction and insoluble fraction.

The soluble fraction obtained was adjusted to pH 4.5 was 35% hydrochloric acid, and centrifuged at 3,000 G for 5 10 minutes to obtain a precipitated fraction. Then, to the precipitated fraction was added water, and the mixture was neutralized with a 20% aqueous sodium hydroxide solution and freeze-dried to obtain a 7S globulin-rich fraction.

On the other hand, 5 fold by weight of ion-exchange 10 water was added to the insoluble fraction obtained by centrifugation at 5,000 G for 10 minutes, and the mixture was extracted by stirring at 22°C for 30 minutes with a propeller while pH of the mixture was maintained at 6.8 with a 20% aqueous sodium hydroxide solution. Then, the 15 mixture was centrifuged at 5,000 G for 10 minutes, and pH of the supernatant obtained was adjusted to 4.5 with 35% hydrochloric acid, followed by centrifugation at 3,000 G for 10 minutes to obtain a precipitated fraction. Then, to the precipitated fraction was added water, and the mixture 20 was neutralized with a 20% aqueous sodium hydroxide solution, and freeze-dried to obtain a 11S globulin-rich fraction.

The ratio of 7S : 11S of each fraction as determined by SDS-polyacrylamide electrophoresis, the purity of 7S 25 globulin (SPE standard) in the 7S globulin-rich fraction

and the purity of 11S globulin (SPE standard) in the 11S globulin-rich fraction, the amount of the chloroform-methanol oil content, the corrected purity of 7S globulin, the corrected purity of 11S globulin, and the content of phytic acid in each fraction are shown in Table 9. Table 10 shows the separation-precipitation rate of the insoluble fraction measured according to the same manner as in Example 1.

Table 9

Composition of each fraction (unit: %)

	Soluble fraction	Insoluble fraction
7S:11S	96:4	13:87
Purity (SPE standard)	93.0	82.9
Crude protein (per dry weight)	95.7	94.2
Chloroform-methanol oil content (per dry weight)	0.65	1.30
Corrected purity	87.0	72.1
Phytic acid (per dry weight)	2.1	2.0

Table 10

Separation-precipitation rate of insoluble fraction

	Without phytase treatment
Separation-precipitation rate ($\mu\text{m/sec}$)	182

10

As seen from the results above, it is clear that the fractions of highly purified 7S and 11S globulins containing little amount of the chloroform-methanol

fraction, or containing little amount of the oil-body-associated protein can be readily obtained by the process of the present invention.

From the results of Example 1 in which the sample
5 solution was immediately cooled without holding the temperature after the heat treatment and from the results in Example 5 in which the temperature was maintained for 20 minutes after heat treatment, it is evident that maintaining the temperature after the heat treatment can
10 improve the separation-precipitation rate.

Example 6

The ionic strength of a 5% curd slurry prepared according to the same manner as in Example 1 was adjusted to 0.14 with sodium chloride, the slurry was stirred at
15 22°C for 30 minutes without adjusting the pH (about pH 4.5) followed by heating to 50°C. The time required for heating was 10 minutes. When the temperature was raised to 50°C, pH of the slurry was adjusted to 5.3 with a 20% aqueous sodium hydroxide solution. Then, phytase (Sumizyme PHY,
20 manufactured by Shin Nihon Chemical Co. Ltd.) was added in a proportion of 0.2% by weight based on the weight of defatted soybeans, and the slurry was stirred for additional 20 minutes with a propeller. Then, the slurry was cooled to 22°C. The time required for cooling was 5
25 minutes. After cooling, the slurry was centrifuged at

5,000 G for 10 minutes to obtain a soluble fraction and an insoluble fraction. The soluble fraction obtained was adjusted to pH 4.5 with 35% hydrochloric acid, followed by centrifugation at 3,000 G for 10 minutes to obtain a precipitated fraction. Then, to the precipitated fraction was added water, and the mixture was neutralized with a 20% aqueous sodium hydroxide solution, and freeze-dried to obtain a 7S globulin-rich fraction.

On the other hand, 5 fold by weight of ion-exchange water was added to the insoluble fraction obtained after centrifugation at 5,000 G for 10 minutes. The mixture was stirred at 22°C with a propeller, and was extracted for 30 minutes while maintaining pH at 6.8 with a 20% aqueous sodium hydroxide solution. Then, the mixture was centrifuged at 5,000 G for 10 minutes, the supernatant obtained was adjusted to pH 4.5 with 35% hydrochloric acid, followed by centrifugation at 3,000 G for 10 minutes to obtain a precipitated fraction. To the precipitated fraction was added water, and the mixture was neutralized with a 20% aqueous sodium hydroxide solution, and freeze-dried to obtain an 11S globulin-rich fraction.

The ratio of 7S : 11S of each fraction as determined by SDS-polyacrylamide electrophoresis, the purity of 7S globulin (SPE standard) in the 7S globulin-rich fraction and the purity of 11S globulin (SPE standard) in the 11S

globulin-rich fraction, the amount of the chloroform-methanol oil content, the corrected purity of 7S globulin, the corrected purity of 11S globulin, and the content of phytic acid in each fraction obtained are shown in Table 11.

5 Table 12 shows the separation-precipitation rate of the insoluble fraction measured according to the same manner as in Example 1.

Table 11

Composition of each fraction (unit: %)

	Soluble fraction	Insoluble fraction
7S:11S	97:3	14:86
Purity (SPE standard)	94.0	81.9
Crude protein (per dry weight)	96.2	92.4
Chloroform-methanol oil content (per dry weight)	0.61	1.24
Corrected purity	88.3	71.7
Phytic acid (per dry weight)	0.05	0.12

Table 12

Separation-precipitation rate of insoluble fraction

	With phytase treatment
Separation-precipitation rate ($\mu\text{m/sec}$)	248

As seen from the results in Examples 5 and 6, it is
10 clear that concomitant use of decomposition of phytic acid with phytase facilitates separation of the soluble fraction and the insoluble fraction and makes it possible to obtain

the 7S globulin-rich fraction and the 11S globulin-rich fraction, both containing less phytic acid.

Industrial Applicability

5 As described hereinabove, according to the present invention, a 7S globulin-containing soluble fraction and a 11S globulin-containing insoluble fraction can be simply and readily fractionated in an industrial scale by combining adjustment of the ionic strength and heat
10 treatment under acidic conditions of a solution containing soybean protein.